267 m μ , $\epsilon = 44136$; $[\alpha]_{D}^{25}$, -2° (C = 0.1 in 0.1 N HCl in methanol). Infrared analysis (KBr pellet) showed two new bands, 5.64 and 5.8 μ , related to carbonyl moieties not present in intact tetracycline.

SUMMARY

Cupric-morpholine complex rapidly inactivates tetracycline in methanol or methanol water solutions. Kanamycin is not affected. This selectivity of inactivation has been used as part of an assay for kanamycin in the presence of tetracycline.

Analytical data of the isolated crystalline reaction product indicate extensive degradation of the tetracycline molecule has occurred. An apparent empirical change from C22H24N2O8 to C17H16NO8 was noted.

REFERENCES

Brockman, W., and Havinga, C., Rec. Trav. Chim., (2) (a) Sakaguchi, T., and Kiyomi, K., J. Pharm. Soc. Japan, 79, (No. 11), 1381(1959); (b) Doluisio, J. T., and Martin, A. N., Jr., "American Chemical Society, Division of Medicinal Chemistry Abstracts," Washington, D. C., meeting, March 1962, p. 7N.
 (3) Grove, D. C., and Randall, W. A., "Assay Methods of Antibiotics," Medical Encyclopedia, Inc., New York, N. Y., 1955 p. 48

Antibiotics, Metical Encyclopedia, Inc., New York, N. Y., 1955, p. 48.
(4) Lamoy, G. V., and Lannon, J. H., "Antibiotics Annual, 1958–1959," Medical Encyclopedia, Inc., New York, N. Y., 1959, p. 790.
(5) Kaplan, M. A., and Buckwalter, F. H., "Antibiotics Annual, 1957–1958," Medical Encyclopedia, Inc., New York, N. Y. J.

N. Y., 1959, p. 507.

Cast Electrode Mount for Self-Stimulation Electrodes By THOMAS J. HALEY, A. M. FLESHER, K. FLYGARE, and R. MYERS

IN A PREVIOUS communication (1), the construc-tion and implementation tion and implantation of self-stimulation electrodes was described. Recently, it was discovered that when stereotaxic equipment other than the Lab-Tronics apparatus was used, the mount would not conform to the skull and loss of both the electrode and the animal resulted. Moreover, the amount of handwork required in producing the Lucite mounts was excessive. These disadvantages have been overcome by casting epoxy resin mounts as described below.

The dimensions in inches of the brass pattern used in preparing the mold for casting the epoxy resin mount are given in Fig. 1. The angles shown are critical since any deviations will result in a mount which will not conform to the skull. The completed brass pattern is shown in Fig. 2A. The mold shown in Fig. 2B is made from a RTV-11 silicone rubber compound (General Electric Co.) mixed according to package directions and poured around the brass pattern in an aluminum form 7/8 in. in diameter and ⁵/₈ in. in depth. This mold sets in 24 hours; several should be prepared. The epoxy mounts are made from Wilhold clear epoxy glue by filling the mold and allowing 4 hours for hardening. All bubbles should be removed by stirring with a fine wire. The mount is removed from the mold by flexing the latter. All excess resin should be removed by sanding with fine emery paper. The completed mount is attached to the rat's skull as previously described (1), and the electrode is inserted and cemented firmly in place.

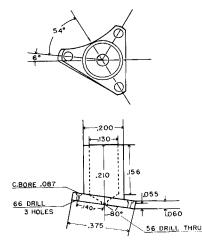


Fig. 1.-Brass pattern of epoxy resin mount with dimensions in inches.

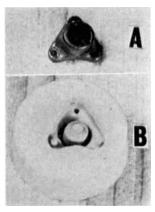


Fig. 2.-Key: A, completed brass pattern; B, completed silicone rubber mold for preparing epoxy resin electrode mounts.

REFERENCE (1) Haley, T. J., et al., THIS JOURNAL, 50, 710(1961).

Received September 17, 1964, from the Laboratory of Nuclear Medicine and Radiation Biology, Department of Biophysics and Nuclear Medicine, School of Medicine, University of California at Los Angeles, Los Angeles. Accepted for publication October 9, 1964. These studies were supported by contract AT(04-1)GEN-12 from the U. S. Atomic Energy Commission, Washington, D. C.